REVIEW ARTICLE

Application of hurdles for extending the shelf life of fresh fruits

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Abstract

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Fruits, when harvested have high moisture content and higher water activity that provides suitable ground for the growth of microorganisms. In addition, the biochemical and metabolic processes do not cease; though the fruit have left their parent plants. The processes such as respiration, senescence, and conversion of starch etc. contribute to the degradation process. Therefore, preservation of these fresh fruits becomes the top most priority for the producers, dealers and vendors. More than 100 methods are followed worldwide for preservation of different kinds and forms of foods. Most of these technologies aim at controlling the microbial growth for food safety, simultaneously altering the composition, colour, and taste of the produce. The likeness and public awareness for consumption of minimally processed or fresh like food has brought the concept of hurdle technology. The technology initially applied to meat preservation, has evolved into preservation of minimally processed foods, processed foods etc. This involves combination of two or more methods that act synergistically, like application of chlorine, ozone, chemical, ultrasound, hyperbaric pressure, active packaging/MAP, edible coating, electromagnetic waves in moderate doses and storage at suboptimal temperature and humidity, causing least damage to the quality parameters, thereby extending the shelf life of fresh fruits.

Keywords: Fruits, climacteric, non-climacteric, hurdles, temperature.

Introduction

Fruits are rich in carbohydrates, vitamins, and minerals and poor in proteins, with pH values ranging from 7.0 to slightly acidic, and exhibit a characteristic high water activity. These conditions make the produce suitable ground for growth of several bacteria, yeasts and moulds (Ramos et al., 2013). Fruits deteriorate rapidly after harvest and in some cases do not reach consumers at optimal quality after transport and marketing. Post harvest, the fruits are still respiring and other metabolic processes still continuing. Even mild bruising can contribute largely to the produce's deterioration through enzymatic reaction and microbial contamination. By the time the fruit reaches the consumer's gut, it goes through different phases, witnessing and sustaining different handling and storage conditions. Spoilage may be caused by a wide range of reactions such as physical, chemical, enzymatic, and microbiological. The various forms of microbiological spoilage are preventable to a large -

degree by a wide range of preservation techniques, most of which act by preventing or inhibiting microbial growth (e.g., chilling, freezing, vacuum packing, atmosphere modified packing and adding preservatives). A smaller number of techniques act by inactivating microorganisms (e.g., pulse electric field, irradiation). Additional techniques restrict the access of microorganisms to products (e.g. packaging). A major trend is that new and emerging preservation techniques, which are coming into use or are under development, include more than one act of inactivation (e.g., electroporation, hypobaric pressure, manothermosonication and addition of bacteriolytic enzymes). A further trend is towards the use of procedures that deliver less heavily preserved products with higher retention of quality, more natural, free from additives, and nutritionally healthier. Less severe preservation procedures are therefore being developed that make use of preservative factors in combinations

to deliver less damage to product quality (Gould, 1996).

Fruit contamination problems can occur in the growing environment. During growth the fruit can become contaminated from sources such as soil, water, animals, birds, and insects. Following production, the processes of harvesting, washing, packaging, and shipping can create additional conditions. The quality of food can be adversely affected by physical, chemical, biochemical and microbiological processes. Quality deterioration caused by microorganisms may include a wide range and types of spoilage that are commercially. The undesirable microbial contamination, however, can adversely affect the fruit quality and hence the final product quality (Lee, 2004).

Food preservation goes long back since the evolution of human beings. As the evolution progressed, so did the means and methods of food preservation. Various methods of control process namely, drying, heating, cooling, salting or pickling, pasteurisation or sterilisation, edible coating, application of MAP, CAP, electromagnetic radiation have been in use for this purpose. With passage of time, consumer's taste and perception of food have been changed, which could be linked to the increased buying power of the consumer and general awareness of the health benefits of fresh produce. With the decline interest in the intermediate moisture food for their saltiness, sugar retention, flavour and colour changes, consumers have started giving emphasis to the freshness of food, with minimum quality deterioration.

The concept of hurdle technology evolved through the idea of keeping the freshness of food intact. Multi-target approach is the essence of hurdle technology, while ensuring product quality by the application of milder but synergistically acting methods (Leistner, 1992). The original Hurdle concept hypothesises the existence of synergy between antimicrobial factors. Building on the idea of food interfering with microbial homeostasis as the hypothesis of food preservation, Leistner developed the hypothesis of the multi-target preservation of foods (Bidlas and Lambert, 2008) as a foundation for the multiple-hurdle concept. Homeostasis as quoted by Leistner and Gorris (1995) "is the constant tendency of microorganisms to maintain the stability and balance of their internal environment such as pH and osmotic strength in the cell". Up to now, more than 50 hurdles are applied for the preservation of foods, which include temperature, relative humidity, pH, water activity, acidity, redox potential, chemicals, antioxidants and preservatives. The food being subjected to milder processing treatments retains the product quality with minimal damage. The storage of high moisture food having high water activity, no longer became a constraint while practicing such mild processing methods. In this method, two or more mild methods of food preservation techniques are applied to the food, so as to create hurdles for the aforementioned spoilage causes. Therein facilitate the increased shelf life of food.

The hurdle technology concept was first applied to keep the freshness of meat (Leistner, 1992; 1994; 2000; Aymerich et al., 2008) and now-a-days its application has been extended to preservation of fresh, minimally processed and processed fruits (Alzamora et al., 1993), vegetables (Niemira et al., 2005), and dairy foods (Del Nobile et al., 2009; Walkling-Ribeiro et al., 2009). The major target of hurdle technology has been the microbial inactivation (Ross et al., 2003). The quantification of different parameters and their synergistic effect on the concept of hurdle technology has been addressed by response surface modelling, gamma and logistic modelling (Bidlas and Lambert, 2008). Application of hurdle technology for minimally processed foods has been reviewed by Alzamora et al. (1993). However, in this article focus will be on the shelf life extension of fresh fruits using hurdle technology.

Classification of fruits

Fruits are most liked food by the consumer all over the world, for their sweet taste, aroma and nutritive value. According to their geographical distribution they can be categorised into tropical, subtropical and temperate fruits.

Tropical fruits: Tropical fruits are grown in tropics and they cannot withstand slight chilling temperature. They require warm climate for fruiting.

Sub-tropical fruits: Subtropical fruits are grown in the sub tropics and Mediterranean's. They require warm or mild temperature but they can withstand mild frost.

Temperate fruits: These fruits are mainly grown in the temperature region (cold-winter) climates and require chilling temperature for the fruiting. They undergo dormancy in winter enabling them to tolerate freezing temperature and low photoperiods. During plant dormancy, visible growth is suspended and all physiological processes are halted or slowed (Luedeling, 2012). Some of the tropical, sub-tropical and temperate fruits are listed in Table 1.

Many of the fruits are seasonal in nature and highly perishable owing to their higher water content. Preserving the qualities of fresh fruits is very tedious as, several biochemical, metabolic, and microbial activities regulate the quality and shelf life of fruits.

They are harvested at full maturity and are selfsufficient to carry out further metabolic activities, such as respiration and ripening, even after being detached from the parent plant. Furthermore, the fruits are again classified into two major categories i.e. climacteric and non climacteric fruits, which have different biochemical and metabolic pathways, in addition to having different chemical composition.

Climacteric fruits: These fruits are characterised by high ethylene biosynthesis, after harvesting from the parent plants, which means the ripening continues even after harvesting. During ripening process of climacteric fruits, the levels of ethylene biosynthetic enzymes 1aminocyclopropane-1-carboxylate (ACC) synthase (ACS) and ACC oxidase (ACO) increases (Yamane *et al.*, 2007). Some of the major fruits of this category are listed in Table 2.

Non-climacteric fruits: The fruits neither synthesise enough ethylene during growth cycle nor does respond to external ethylene stimulus. So these fruits must be harvested from the parent plant when fully matured. Harvesting the fruits before they are fully matured will cause the uneven growth and quality degradation. The metabolic rates of these fruits are slower compared to the climacteric fruits (Prasanna *et al.*, 2007). Most berries and citrus fruits come under this category (Table 2).

For both climacteric and non climacteric fruits, the ripening process is hastened by the addition of ethylene; nevertheless the ripening process can be delayed by flushing out naturally evolved ethylene through aeration.

Food spoilage causes

The fresh fruit can be spoiled by different factor i.e. microbial growth, biochemical changes such as ripening and respiration, caused by enzymatic activities. The major reasons of spoilage during storage could be attributed to the intrinsic properties of food, contamination during harvesting, handling and processing, mechanical wounds inflicting to enzymatic activity along with temperature abuse.

Microbial spoilage: Fruits can be contaminated with a wide range of microorganisms during harvesting, handling, transportation, and processing. Certain microflora are already present in the fruits prior to harvesting. Some of the microbes grow and spoil the produce during subsequent storage and distribution. The type of biochemical reactions followed or the microbial growth depend on the environmental factors and intrinsic properties of produce. For the past few decades public health concern has been largely attributed to contaminated fruits and vegetables. The different pathogens most frequently linked to fruit and vegetable produce-related outbreaks generally include bacteria such as *Escherichia coli O157: H7, Salmonella spp.* and *Listeria spp.* Many pathogens and bacteria including *Bacillus, Salmonella, Listeria, Staphylococcus,* and *Escherichia* are capable of adhering to and forming a biofilm on fruits and vegetable surfaces (Bilek and Turantaş, 2013).

Fruit ripening and quality degradation: Ripening is the one of the most complex plant metabolic pathway. The fruits undergo various changes i.e. translocation of nutrients, conversion of starch into sugar, phenolic compound development, increased respiration, chlorophyll degradation/ carotenoid and anthocyanin synthesis, degradation of flavanoids, tannins, pectin degradation, cell wall softening and most importantly ethylene biosynthesis. All these processes bring about desirable changes in texture, colour and aroma of the fruit. Most of these metabolic activities are governed by a plant hormone (Prasanna et al., 2007) ethylene. Ethylene regulates every stages of plant growth, from sprouting of seed to leaf development and from flowering to fruit maturity and ripening. These hormones are produced when triggered by wound, temperature or water stress, wind velocity etc. Ethylene expedite the ripening and respiration processes, causing senescence and cellular disintegration, hydrolysis of compounds, resulting in quality degradation, thus limiting the shelf life of the fruit. The ripening process can be divided into pre-climacteric and climacteric stages. During the pre-climacteric stage the ethylene production is slow, thus delaying the maturity and so does the respiration rate. During the climacteric stage, however, respiration rate increases and there is higher ethylene production in the plant cell.

Apart from quality degradation due to ripening, fruits suffer from chilling injury while in cold storage. In many cases the fruits are transported and stored below their recommended temperature and relative humidity. This result in browning of skin and pulp, leading to quality deterioration and eventually decrease in the market price of the fruits.

Preservation of fresh fruits

Inhibition of microbial load: Fresh fruit surface could be a host of various virus, bacteria, yeast and mold which are manifested at any phases of pre harvest and post harvest conditions. Microbial load of the fresh fruits or minimally processed fruits can be reduced by application of several non- thermal technologies such as pulsed electric fields (PEFs), ionizing radiation and-

| Tropical | Sub-tropical | Temperate | |
|-------------------|---------------|--------------|--|
| Acerola | Avocado | Apple | |
| Banana | Fig | Apricot | |
| Cashew apple | Dates | Blackberry | |
| Cherimoya | Grapes | Blackcurrant | |
| Custard Apple | Grape fruit | Blueberry | |
| Dragon fruit | Lime | Cherry | |
| Durian | Litchi | Gooseberry | |
| Elephant apple | Olive | Grapes | |
| Guava | Orange | Kiwi Fruit | |
| Indian gooseberry | Mandarin | Nectarine | |
| Jackfruit | Passion Fruit | Mulberry | |
| Mango | Pomegranate | Peach | |
| Papaya | Loquat | Pear | |
| Pineapple | Longan | Melon | |
| Sapote | - | Plum | |
| Star apple | | Quince | |
| Sweetsop | | Raspberry | |
| Water melon | | Strawberry | |

Table 1: Classification of fruits based on geographical distribution

Table 2: Classification of fruits based on ethylene biosynthesis

| Climacteric fruits | Non-Climacteric fruits | | |
|--------------------|------------------------|--|--|
| Apples | Blackcurrant | | |
| Apricots | Blueberry | | |
| Avocados | Carambola (star fruit) | | |
| Bananas | Cashew apple | | |
| Custard Apple | Cherry | | |
| Date | Cucumber | | |
| Figs | Grape | | |
| Guavas | Grapefruit | | |
| Honeydew melon | Lemon | | |
| Jack fruit | Litchi | | |
| Kiwi fruit | Lime | | |
| Mangoes | Longan | | |
| Musk melon | Loquat | | |
| Nectarines | Mandarin | | |
| Papaya | Melon | | |
| Passion fruit | Pineapple | | |
| Peaches | Pomegranate | | |
| Pears | Olive | | |
| Persimmons | Orange | | |
| Plum | Raspberry | | |
| Sapote | Strawberry | | |

ultrasonication at ambient or sub-lethal temperatures. Many of these processes require very high treatment intensities; however, to achieve adequate microbial destruction in fresh fruits, combining non-thermal processes with conventional preservation methods significantly enhances their antimicrobial effect, so that lower process intensities can be used (Ross et al., 2003). For conventional preservation treatments, optimal microbial control is achieved through the hurdle concept, with synergistic effects resulting from different components of the microbial cell being targeted simultaneously with irradiation, radio frequency heating, microwave blanching, and O₃ treatment, at a lower level, so as to keep the quality degradation in the process in the form of loss of colour, texture and other essential nutrient to the minimum. The aforementioned processes can be coupled with application of essential oils, chemicals, cold storage/ refrigerated storage, freezing, aseptic packaging, edible coating, MAP, CAP, active packaging, pulsed UV light, irradiation, radio frequency heating, chlorine water etc.

Inhibition of respiration loss: Fresh fruits and vegetables continue to respire, even though detached from the parent plant, resulting in weight and moisture loss. In the respiration process, the glucose is oxidised to form CO₂ and moisture. Creating an oxygen deficit environment by the application of CAP or MAP would greatly reduce the respiration rate and thereby the moisture loss. In MAP, the storage environment is modified by allowing CO₂ to build up, ultimately restricting the availability of O_2 to the plant tissue. In CAP, however, constant CO₂ and O₂ ratio is maintained by purging CO₂ into the storage environment. The respiration rate can be decreased by storing the fresh produce at lower temperature, using MAP, CAP or active packaging methods. Though respiration rate can be lowered using such technologies, the water accumulation resulting from respiration can be a cause of microbial growth. This problem can be solved by using one or more desiccants in combination, in the MAP and CAPs (Mahajan et al., 2008).

Inhibition of ethylene biosynthesis: Ethylene biosynthesis can be controlled by maintaining low temperature, creating an oxygen deficit environment, increasing CO_2 concentration, blocking the ethylene generating sites by introduction of exogenous ethylene, applying ethylene biosynthesis inhibitors like vinyl glycine analogs, 1-methylcyclopropene (1-MCP), hydroxylamine analogs, Ca^{2+} , orthophosphate ions, cobalt chloride (CoCl₂), decouplers and membrane disruptive agents like 2,4-Dinitrophenol (DNP) and

carbonyl cyanide m-chlorophenylhydrazone (CCCP) in small doses, free radical inhibitors such as n-propyl gallate and sodium benzoate and polyamines (Apelbaum *et al.*, 1981; Yang and Hoffman, 1984).

Potential hurdles for fresh whole fruits preservation

In order to achieve the safe storability of the fresh fruits, the hurdle technology must comply with the multi-target methods. Post harvest, the fresh produce should be attended to ensuring the safety of the stored products. The fruits should be washed and sanitized properly prior to storage or packaging. Even hot water immersing, rinsing and brushing can decrease the microbial load to a significant level (Fallik, 2004). Some of the commonly used decontaminating and sanitizing agents are detergent products, chlorine, electrolyzed water, aqueous chlorine dioxide and acidified sodium chlorite, aqueous ozone, peroxyacetic acid, hydrogen peroxide, organic acids, alkaline products and iodine (Sapers, 2009). A few sanitizing agents, however, impart off flavour, colour and texture to the fresh fruits. Several antimicrobial washing solutions, O₃, UV–C radiation, super high O₂, hexanal, N₂O and noble gases alone or in combination, are presently considered as promising treatments (Utto et al., 2008; Artés et al., 2009).

Chlorine wash: The use of chlorine as a produce disinfectant is very common in the food industries. It is generally used in the following forms: chlorine gas, calcium hypochlorite and sodium hypochlorite and there has been much research into the efficacy of chlorine as a sanitizer for produce decontamination. There are public issues however, regarding the residual effect of chlorine on the fresh fruits and vegetable surfaces. Moreover, chlorine is not efficient in eliminating the biofilm forming bacteria. Goodburn and Wallace (2013) emphasized the use of chlorine as decontamination methods after reviewing various technologies for enhancing the storage lie of fresh produce. In some cases chlorine wash was found to be more effective against fungal infection than ozone (Crowe et al., 2012). Shin et al. (2012) when treated strawberries with 50 ppm aqueous chlorine dioxide and 5 kJ/m² ultraviolet-C irradiation and packed with rice bran protein film containing 1% grapefruit seed extract, observed a better microbial control. Chlorine is a common efficient sanitation agent but there is the risk of undesirable by-products upon reaction with organic matter and this may lead to new regulatory restrictions in the future. For example application of chlorine for decontamination of strawberries can result in off flavour development and anthocyanin degradation.

Moreover; its efficacy is poor for some products. Similarly application of H_2O_2 as sanitizer is well documented.

peroxide treatment: Hydrogen Hvdrogen peroxide is an effective sanitizing agent and has been approved for food use. The oxidative as well sanitizing properties makes it an ideal choice for use in food industry. The effectiveness of this sanitizer has been documented for different blueberries, strawberries, lemon, and melon among others (Ukuku, 2004; Alexandre et al., 2012; Cerioni et al., 2013; Li and Wu, 2013). Li and Wu (2013) have studied the efficacy of this sanitizer in combination to retard the growth of Salmonella with other chemicals and recommended 0.5 mg/ml acetic acid plus 5000 ppm Sodiumdisulphide (SDS), 200 ppm hydrogen peroxide in combination with 5000 ppm SDS as an alternative to the use of chlorine-based washing solution for blueberries. Hydrogen peroxide treatment, however, imparts anthocyanin degradation in the pericarp of litchi (Ruenroengklin et al., 2009).

Application of chemicals: Application of chemicals like 1-methylcyclopropane (1-MCP), oxalic acid, inhibits chilling injury. 1-Methylcyclopropene (1-MCP) is an inhibitor of ethylene receptors and delays ripening of horticultural products. The use of 1-MCP is a potentially useful tool for commercial application to reduce the ripening process, senescence, retard fruit softening, alleviate chilling injury, maintain quality, and extend shelf life of fruit, vegetables, and ornamental species (Prasanna et al., 2007). 1-MCP treatment is now accepted for application as a ripening delaying agent. Inappropriate dosing might induce uneven colouring of the pericarp of fruits. Similarly application of oxalic acid, salicylic acid, methyl jasmonate and nitric oxide alleviates the chilling injury symptoms in various fruits like pomegranate (Sayyari et al., 2009), loquat (Cao et al., 2010), plum (Luo et al., 2011), tomato (Zhao et al., 2011), cucumber (Yang et al., 2011), mango (Li et al., 2014).

Ozone treatment: Ozone is a highly effective sanitizing agent. Application of both aqueous and gaseous ozone has been in use for decontaminating meat, poultry, fish, fruits and vegetables (Najafi and Khodaparast, 2009). Ozone has been known to reduce sprouting in potatoes and disinfesting tomatoes. During storage, if the temperature is not maintained, bacterial growth can resurface (Kim *et al.*, 2003; Tzortzakis *et al.*, 2008). Since ozone is an oxidising agent, it may impair off flavour or colour disintegration in anthocyanin containing fruits. Ozone could be seen as an alternative to refrigeration in order to enhance

tomato shelf life in areas where cold facilities are not available. Zambre *et al.* (2010) has observed that shelf life of tomatoes can be enhanced by 12 days when ozone treated tomatoes were stored at 15°C. Ozone application has improved the antioxidant capacity of papaya fruit and it was observed that the sweetness and overall acceptability was better with ozone treated fruits. Thus ozone can be used as a non-thermal and safe preservation technique for papaya fruit (Ali *et al.*, 2014). Overexposure of ozone may lead to off flavour development and environmental pollution. It may not be as effective as chlorine on certain fruits (Crowe *et al.*, 2012).

Irradiation: For the marketing of raw or minimally processed foods, cold decontamination process is a requirement for effective management and control of the quality in the food chain. Irradiation is such a control measure in the production of several types of raw or minimally processed foods such as poultry, meat and meat products, fish, seafood, and fruits and vegetables. This technology ensures complete product safety from the vegetative form of pathogens as well as parasites and is aptly termed as cold pasteurisation. Irradiation is a safe technology and the critical limits of the doses are well established and can be corrected if necessary and have been recognized as such by the FAO/WHO Codex Alimentarius Commission. It certainly merits the attention of industry and public health authorities (Molins et al., 2001). Research on irradiation of several tropical fruits such as papayas, mangoes, litchi showed that the chemical, sensory and nutrient qualities of these fruits were well retained at 1.0 kGy, and the fruits would ripen normally or slightly delayed (Moy, 1003). Irradiation does of (0.3-0.7 kGy) reduced post harvest decay in mango. Higher gamma radiation dose (6-10 kGy), however, imparted radiation injury on the fruits like mango (Mahto and Das, 2013).

High voltage electric field: Existence of electrostatic force of repulsion has been first observed by Niccolo Cabeo in 1629. When sufficiently high electric field (AC or DC), in the order of kilovolts, in the domestic or industrial frequency range (50 or 60 Hz) is applied across the food, which is composed of complex molecules like carbohydrate, protein, fat, vitamins, polyglycerides and water, it polarizes the bipolar molecules (Mohapatra and Mishra, 2011). This phenomenon interferes with the metabolic pathways of fresh produce when treated with high voltage electric field (HVEF). Few studies have reported on the effect of HVEF on shelf-life of food materials. Toda (1990) (cited by Palanimuthu et al., 2009) treated lettuce, spinach and komatsuna with HVEF and observed a reduction in the respiration rates. Similarly results were

reported by Kharel *et al.* (1996) for HVEF treated pear, plum, banana, apple and sweet pepper. Atungulu *et al.* (2004) observed the reduced rates of respiration in apple under HVEF. Bajgai *et al.* (2006) treated Emblic fruit (Phyllanthus emblica L.) with HYVF (430 kV/m) for 2 h and concluded that HVEF treated fruits have better freshness compared to untreated ones. Palanimuthu *et al.* (2009) observed that application of HVEF (2-8 KV/cm) reduced the respiration rate of cranberry and the weight loss was in the range of 23.2– 30.4% after 3 weeks of storage. Most reports on HVEF application in food has concentrated on fluid foods; its application on solid fruits is limiting. Thus commercialization of this process would require much more information; therefore, research work.

Electromagnetic wave application: Decontamination using organic solutions and disinfectants can ensure the safety from the spoilage organisms clinging to the pericarp of the fruits, but some insects and viruses cannot be detected as they grow from the embryo or may be in dormant stage. Outwardly invisible, they may grow and spoil the fruits during storage if a homeostatics situation arises. In that case irradiation or electromagnetic wave propagation like microwave (MW), radiofrequency (RF) through the fresh fruits will be beneficial. Since the electromagnetic waves penetrate to a greater depth and result in volumetric heating. Mild heat treatment, such as MW, RF heating along with other decontamination treatment has a great potential in fresh fruit marketing (Ikediala et al., 2000; Zhang et al., 2006; Birla et al., 2004). Pulse UV (PUV) light also found to be effective in reducing microbial load for an exposure period of 10 s on a varieties of fruits including apples, kiwi, lemon, nectarines, oranges, peaches, pears, raspberries, and grapes (Lagunas-Solaret al., 2006). They have recommended PUV technique for commercial scale disinfection measures as non-chemical method; however, for maximum disinfection efficiency, coherent PUV sources must be combined with dispersing reflectors, and fruits must be handled to ensure uniform exposure to multidirectional incident beams. Application of UV-C treatment also alleviates the chilling injuries in fruits (Pongprasert et al., 2011). UV-C treatment has potential to delay postharvest fruit senescence and especially control decay in different fruit and vegetable species (Maharaj et al., 1999; Barka et al., 2000; Erkan et al., 2001; Marguenie et al., 2003; Allende and Artes, 2003; Allende et al., 2006). The exposure to UV-C delays fruit softening which is one of the main factors determining fruit postharvest life (Pan et al., 2004). Barka et al. (2000) found that UV-C decreased the activity of enzymes involved in tomato cell wall degradation and delayed the fruit softening.

Reduction of strawberry fruit softening, less decay, and increase in phenoloc content by UV-C application has also been reported (Baka *et al.*, 1999; Erkan *et al.*, 2008).

Ultrasound application: Ultrasound is one of the newest non-thermal methods to extend shelf life of fresh fruits during storage. It is perceived as safer, nontoxic, environmental friendly process without any detrimental effect on human health. The effectiveness of ultrasound depends on wave frequency, power and treatment time. Sound waves carry acoustic energy and can be transmitted though pressure fluctuations in air, water or any other elastic media. These acoustic waves when encounter any deviation of particles from their mean position, they try to level it off; thereby passing some amount of energy to the next particle. So the disturbances go on in a cyclic manner, forming compression, through increase in pressure and rarefaction, though decrease in pressure, in the medium. Sound waves can be classified into three categories i.e. supersonic (frequency< 20 Hz), audible (20 Hz <frequency>20 kHz), or ultrasound (frequency>20 kHz). Ultrasound waves can again be classified into two categories, high frequency-low energy waves that are used for non destructive quality measurement and analysis and low frequency -high energy waves or power ultrasound. Power ultrasound usually refers to the frequency range between 20-40 kHz (Mohapatra and Mishra, 2011). Numerous studies have been attempted to explain the effect of ultrasound on fruits, vegetables (Bilek and Turantas, 2013). Aday et al. (2013) demonstrated the effect of ultrasound on strawberry quality and concluded that ultrasound power levels of 30-60W for 5-10 min treatment time has resulted in improved quality and can be used to enhance the shelf life of the product. Ultrasound alone is not effective against microorganism; hence its application in combination with other treatments such as heat, pressure and chemical treatment will enhance its efficiency.

Hyperbaric pressure treatment: Hyperbaric treatment is a physical postharvest preservation technique in which fresh produce are subjected to an elevated pressure environment ranging from 0.1 to 1.0 MPa (Goyette *et al.*, 2012). Recently, a few studies have shown that hyperbaric treatment provides beneficial effects on extending the shelf-life of some fruits and vegetables such as sweet cherries, peach, strawberries, mume fruit, apple, tomato, mango and Japanese pear among others. Baba *et al.* (2003) showed that shelf life of mume fruit subjected to 0.5 MPa for 5 days was prolonged through suppression of respiratory CO_2 and ethylene production. It was also reported that

pressure treatment could maintain a commercially acceptable color quality, reduce weight loss, and protect against chilling injuries. Romanazzi et al. (2001, 2008) studied the effect of short hyperbaric treatments on postharvest decay of sweet cherries, strawberries, and table grapes and found that the incidence of brown rot, gray and blue mold, and total rot was greatly reduced after storage at 20°C. Hyperbaric pressure treatment (0.3 to 0.9 Mpa) of tomatoes, during storage could reduce respiration rate and maintain freshness and quality attributes of tomato fruit, with enhanced lycopene content was at the end of ripening period (Goyette, 2010; Goyette et al., 2012; Liplap et al., 2013a,b). A hypobaric pressure (50 kPa, 4 h) treatment of strawberries reduced the fungal rot (Hashmi et al., 2013a) and the cause of delayed decay was attributed due to stimulation of defence-related enzymes (Hashmi et al., 2013b). Hypobaric pressure treatment has potential as an alternative non-chemical postharvest disinfestation method for fresh fruits; the cost factor however prohibits traders to adopt it commercially.

packaging-MAP/CAP/CAS: Active On decontamination, the fresh produce can be further stored in active packaging system. Active packaging is a new concept that has arisen as a response to continuous consumer demand and market trend. This technique concerns with the substances that adsorb/absorb CO2, O2, flavour/odours, moisture, ethylene and those microbes that release CO_2 , antimicrobial agents and antioxidants (Vermeiren et al., 1999). The O₂, CO₂, ethylene scavengers and microbial agents are placed inside the packaging system, so as to effectively improve the packaging environment. The essential oil extracted from spices and plants have anti fungal and anti microbial properties, those can be suitably used in active packaging system without impairing any off odour to the product. Plasticized protein coating on polypropylene films works as antimicrobial agent when incorporated with nisin and whey protein isolate, the films with bacteriocins absorber can be used as an active packaging film (Scannell et al., 2000; Lee et al., 2008). High or low level of O₂ concentration or passive modified atmospheric packaging system is usually adopted in these cases. However, the high O₂ may induce oxidative reaction, resulting in product quality deterioration (Zheng et al., 2007). They recommended to optimize the O_2 concentration for each produce. MAP when combined with other pre-treatments such 1-MCP, essential oil enhances the shelf life of litchi, sweet cherry, table grapes (Sivakumar et al., 2008, De Reuck et al., 2009; Sivakumar and Korsten, 2006: 2010; Serrano et al., 2008). Active, MAP or CAP too have their fallouts. Disbalance in the atmospheric condition could lead to fermentation and off flavour development in the fruits.

Edible film: Edible coating are now been extensively used in storing both processed, minimally processed and fresh whole fruits and vegetables. The edible coatings are chosen keeping in view of the future needs, such as in case of active packaging system where, the film material can react with the food component and prevents undesirable changes. The edible coating can also be used to deliver certain bioactive compounds into the food system. The edible films could be based on fruit (apple puree), vegetable starch (pumpkin), fruit wax, gum cordia, gum Arabic, pectin, carboxymethyl cellulose, chitosan, algenate, whey protein isolate aloe-vera gel with antimicrobial properties (Martínez-Romero et al., 2006; Rojas-Graü et al., 2007; Sothornvit and Pitak, 2007; Sothornvit and Rodsamran, 2008, Saucedo-Pompa et al., 2009; Hag et al., 2013; Lago-Vanzela et al., 2013; Arnon et al., 2014). The edible films have poor mechanical and barrier properties when compared to synthetic polymers, which have lead to the reinforcement of nanocomposites in to biopolymers for improving their properties and enhancing their cost-price-efficiency. However, there are many safety concerns about nanomaterials, as their size may allow them to penetrate into cells and eventually remain in the human organism. While the properties and safety of the materials in their bulk form are usually well known, the nano-sized counterparts frequently exhibit different properties from those found at the macroscale, and there is limited scientific data about their eventual toxicological effects. So the need for accurate information on the effects of nanomaterials on human health following chronic exposure is imperative before any nanostructured food packaging is available for commercialization (Falguera et al., 2011).

Essential oils and phenolics: Plant essential oils and phenolics are antioxidants and their use in Indian cuisine is centuries old. With growing awareness on Indian food and their beneficial effect on health has promoted the use of spices in the food system. Of late significant research is being conducted on the spices and the essential oils derived from them like vanillin. For their antioxidant properties, the essential oils derived from spices and other plant phenolic compounds can be used as coating or in the packaging film ingredient for enhancing storage life as well as to adsorb off orders. These compounds have been included in the list of generally recognized as safe (GRAS) compounds by FDA (Serrano *et al.*, 2005). Though these plant derivatives are regarded as safe and

can be used in place of chemical agents, their commercial viability is yet to be chalked out.

Bacteriocins: Bacteriocins are basically proteinaceous toxic compounds, produced by different groups of bacteria to inhibit the growth of similar or closely related bacterial strain. Many lactic acid bacteria (LAB) produce bacteriocins with rather broad spectra of inhibition. Several LAB bacteriocins offer potential applications in food preservation, and the use of bacteriocins in the food industry can help to reduce the addition of chemical preservatives as well as the intensity of heat treatments, resulting in foods which are more naturally preserved and richer in organoleptic and nutritional properties. In addition to the available commercial preparations of nisin and pediocin PA-1/AcH, other bacteriocins (like for example lacticin 3147, enterocin AS-48 or variacin) also offer promising perspectives. Broad-spectrum bacteriocins present wider uses, while narrow-spectrum potential bacteriocins can be used more specifically to selectively inhibit certain high-risk bacteria in foods like Listeria monocytogenes without affecting harmless microbiota. Bacteriocins can be added to foods in the of concentrated form preparations as food preservatives, shelf-life extenders, additives or ingredients, or they can be produced in situ by bacteriocinogenic starters, adjunct or protective cultures. Immobilized bacteriocins can also find application for development of bioactive food packaging. In recent years, application of bacteriocins as part of hurdle technology has gained great attention. Several bacteriocins show synergistic effects when used in combination with other antimicrobial agents, including chemical preservatives, natural phenolic compounds, as well as other antimicrobial proteins. This, as well as the combined use of different bacteriocins may also be an attractive approach to avoid development of resistant strains. The effectiveness of bacteriocins is often dictated by environmental factors like pH, temperature, food composition and structure, as well as the food microbes (Gálvez et al., 2007). Martínez-Castellanos et al. (2011) demonstrated the quality preservation of litchi bv application of Lactobacillus plantarum. Bacteriocins can be the used in active packaging films or in edible coating for the fresh whole fruits. Cao et al. (2011) has observed the synergistic effect of benzothiadiazole-7-carbothioic acid S-methyl ester (BTH) on biocontrol agent Pichiamembranefaciens in a controlling postharvest blue mould decay in peach fruit. Liu et al. (2013) has reviewed the potential of lactic acid bacteria as biocontrol agent against the pathogenic bacteria, responsible for fruit decay. Some pathogenic bacteria develop bacteriocin resistance;

moreover the dynamics of bacteriocin action depends on the type of fruit and microflora, thereby limiting its efficacy.

Storage temperature: It is well known fact that temperature has a great say whilst several preservation technology has been developed for food preservation.

Application of heat during blanching/pasteurisation or removal of heat during refrigeration has long been practiced in the food industry. In case of fresh fruits, storage temperature is of utmost importance as this parameter decides the viability and growth probability of microorganisms. Since microbes require optimal temperature and water activity for survival and growth, storage at suboptimal temperature conditions enhances the product storage life. As previously discussed, the biochemical reactions affecting the fresh fruits quality also require optimal temperature for the post harvest changes; can be affected or in some cases delayed under low temperature storage, which has been demonstrated by various researchers for fresh fruits such as pears (Villalobos et al., 2011), banana (Khanbarad et al., 2012), pineapple (Hong et al., 2013). Some fruits however, are prone to chilling injuries, marked by browning of the skin and pulp and loss of flavour. In many cases fruits are stored and transported below the optimal temperature, which results in quality deterioration.

Storage relative humidity: When fruits are stored under higher relative humidity, it affects the desiccation and shrivelling of peels of the fruits and in some cases improves the shelf life, as such high relative humidity also favours growth of fungus and molds, therefore should be combined with low temperature storage. Sharkey and Peggie (1984) had observed that the shelf life of cherries and lemons was extended when they were stored under high relative humidity (95-99 %). in most citations relative humidity is combined with temperature of storage and does not have pronounced effect on the fruit quality (Shin *et al.*, 2008). Recommended temperature and relative humidity of storage for different fruits (FAO, 2013) are presented in Table 3.

Pre-cooling: Pre-cooling fruits and vegetable to remove field heat has been in practice since long. It reduces the metabolic rate and improves the shelf life of commodity. Tatsuki *et al.* (2011) applied pre-cooling for 24 h at -1 or -3° C followed by 1 MCP treatment to Tsugaru apple. The fruits were observed for their ethylene production, firmness, and acidity. The pre-cooled fruits maintained high acidity and firmness with low level of ethylene synthesis.

| Fruit | Storage (°C) | Temperature | Storage (%) | Relative | Humidity | Storage (days) | life |
|-----------------------|-----------------|-------------|----------------|----------|----------|----------------|------|
| Apple | -1-4 | | 90-95 | | | 30-180 | |
| Apricot | -0.5-0 | | 90-95 | | | 7-21 | |
| Asian pear | 1 | | 90-95 | | | 150-180 | |
| Atemoya | 13 | | 85-90 | | | 28-42 | |
| Avocado | 3-13 | | 85-90 | | | 14-56 | |
| Babaco | 7 | | 85-90 | | | 7-21 | |
| Banana /Plantain | 13-15 | | 90-95 | | | 7-28 | |
| Barbados cherry | 0 | | 85-90 | | | 49-56 | |
| Blackberry | -0.5-0 | | 90-95 | | | 2-3 | |
| Black sapote | 13-15 | | 85-90 | | | 14-21 | |
| Blueberries | -0.5-0 | | 90-95 | | | 14-21 | |
| Breadfruit | 13-15 | | 85-90 | | | 14-42 | |
| Caimito | 3 | | 83-90 90 | | | 21 | |
| Calamondin | 9-10 | | 90 90 | | | 14 | |
| | | | | | | | |
| Cantalupo | 0-2 | | 95 85 00 | | | 5-15 | |
| Carambola | 9-10 | | 85-90 | | | 21-28 | |
| Cashew apple | 0-2 | | 85-90 | | | 35 | |
| Chayote | 7 | | 85-90 | | | 28-42 | |
| Cherimoya | 13 | | 90-95 | | | 14-28 | |
| Cherries | -1-0.5 | | 90-95 | | | 14-21 | |
| Chicory | 0 | | 95-100 | | | 14-21 | |
| Coconut | 0-1.5 | | 80-85 | | | 30-60 | |
| Cranberries | 2-4 | | 90-95 | | | 60-120 | |
| Cucumber | 10-13 | | 95 | | | 10-14 | |
| Currants | -0,5-0 | | 90-95 | | | 7-28 | |
| Custard apple | 5-7 | | 85-90 | | | 28-42 | |
| Dates | -18-0 | | 75 | | | 180-360 | |
| Durian | 4-6 | | 85-90 | | | 42-56 | |
| Feijoa | 5-10 | | 90 | | | 14-21 | |
| Fig | -0.5-0 | | 85-90 | | | 7-10 | |
| Grape | -0.5-0 | | 90-95 | | | 14-56 | |
| Grapefruit | 10-15 | | 85-90 | | | 42-56 | |
| Guanabana | 13 | | 85-90 | | | 7-14 | |
| Guava | 5-10 | | 90 | | | 14-21 | |
| Jaboticaba | 13-15 | | 90-95 | | | 2-3 | |
| Jackfruit | 13 | | 85-90 | | | 14-42 | |
| Kiwano | 10-15 | | 90 | | | 180 | |
| Kiwifruit | -0.5-0 | | 90-95 | | | 90-150 | |
| Kumquat | 4 | | 90-95 | | | 14-28 | |
| Lemon | 10-13 | | 85-90 | | | 30-180 | |
| Lime | 9-10 | | 85-90 | | | 42-56 | |
| Longan | 1-2 | | 90-95 | | | 21-35 | |
| Loquat | 0 | | 90 | | | 21 55 | |
| Lychee | 1-2 | | 90-95 | | | 21-35 | |
| Mamey | 13-18 | | 85-95 | | | 14-42 | |
| Mandarin | 4-7 | | 90-95 | | | 14-42 | |
| Mango | 13 | | 90-93 90-95 | | | 14-28 | |
| Mangosteen | 13 | | 90-93 85-90 | | | 14-21 | |
| | 7-10 | | 83-90 90-95 | | | | |
| Melon | | | | | | 12-21 | |
| Nectarine | -0.5-0 | | 90-95 | | | 14-28 | |
| Olives, fresh | 5-10 | | 85-90 | | | 28-42 | |
| Orange | 0-9 | | 85-90 | | | 56-84 | |
| Papaya | 7-13 | | 85-90 | | | 7-21 | |
| Passionfruit Peach | 7-10 -0.5-0 | | 85-90 90-95 | | | 21-35 14-28 | |
| | | | | | | | |

Table 3: Recommended temperature and relative humidity for fruits and the approximate storage life under these conditions.

| _ | | | |
|---------------|----------|-------|--------|
| Pear | -1.5-0.5 | 90-95 | 60-210 |
| Cucumber | 5-10 | 95 | 28 |
| Persimmon | -1 | 90 | 90-120 |
| Pineapple | 7-13 | 85-90 | 14-28 |
| Pitaya | 6-8 | 85-95 | 14-21 |
| Plum | -0.5-0 | 90-95 | 14-35 |
| Pomegranate | 5 | 90-95 | 60-90 |
| Prickly pear | 2-4 | 90-95 | 21 |
| Quince | -0.5-0 | 90 | 60-90 |
| Rambutan | 10-12 | 90-95 | 7-21 |
| Raspberries | -0.5-0 | 90-95 | 2-3 |
| Sapodillla | 15-20 | 85-90 | 14-21 |
| Strawberry | 0-0.5 | 90-95 | 5-7 |
| Tart cherries | 0 | 90-95 | 3-7 |
| Tomato (MG) | 12.5-15 | 90-95 | 14-21 |
| Tomato (red) | 8-10 | 90-95 | 8-10 |
| Tree tomato | 3-4 | 85-90 | 21-28 |
| Watermelon | 10-15 | 90 | 14-21 |
| White sapote | 19-21 | 85-90 | 14-21 |
| Yellow sapote | 13-15 | 85-90 | 21 |
| (0 EAO 2012) | | | |

(Source: FAO, 2013)

Table 4: Hurdles applied to different fresh fruits for extension of shelf life

| Fruits | Treatments | Shelf life extension/microbial inactivation | References |
|------------|--|---|---|
| Apple | gamma irradiation (200–400 Gy)+ biocontrol agent (Pseudomonas fluorescens) | Improved quality in 3 months storage period than control | Mostafavi <i>et al.</i> , 2013 |
| | Harvest maturity (190 days after blossoming)+ low (0- 1°C temperature) + CA (1–2 kPa O2/<1 kPa CO2) | Less flesh browning after 2 months storage | Kweon <i>et al.</i> , 2013 |
| Avocado | 1-MCP+ waxing | Delayed ripening after 19days | Jeong <i>et al.</i> , 2003 |
| Banana | 1-MCP (0.5μl/l)+ polyethylene bagging 1-MCP (1000 nL/ L for 4 h at 25 °C)+ non-perforated PE (MAP) | 58 days 100 days | Jiang <i>et al.</i> , 1999 Ketsa <i>et al.</i> , 2013 |
| Blueberry | 100 mg/L chlorine or 1 mg/L aqueous ozone+air blast freezing (60 s), storage at 18°C | 12 months | Crowe <i>et al.</i> , 2012 |
| Cherry | Essential oil+ MAP | Delayed decay after 16 days storage | Serrano <i>et al.</i> , 2005 |
| | Aleovera coating+ low temperature (1°C, 96 %RH) | Less loss of quality | Martínez- Romero <i>et al.</i> , 2006 |
| | Hexanal vapour +1 MCP | 30days | Sharma <i>et al.</i> , 2010 |
| Carambola | MAP +low temperature (10°C) | Delayed solubilization and loss of firmness after 21 days | Ali et al., 2004 |
| Dates | UV-C and neutral electrolyzed water (pH 7.2, ORP 814 mV, and 300 mg L^{-1} of free chlorine) | 30 days storage life | Jemni <i>et al.</i> , 2014 |
| Grape | MAP+essential oil | 56 days | Valero <i>et al.</i> , 2006 |
| Kiwi fruit | MAP+coating with sodium algenate | 13 days | Mastromatteo <i>et al.</i> , 2011 |
| Lemon | H ₂ O ₂ +hotwater+ hydrogen peroxide followed by potassium phosphate | Control of mold and rot | Cerioni <i>et al</i> , 2013 |

| Litchi | sodium hypochlorite, potassium metabisulfite, hydrochloric acid and ascorbic acid dip+gamma does 92.4 kGy) | 30-45 days | Kumar <i>et al.</i> , 2012 |
|--------------------|---|--|---|
| | 1% NaCl+2% wax coating+1 Kgy radiation dose | 24 days | Pandey <i>et al.</i> , 2013 |
| Mango | Dipping in hot 0.5% Na ₂ S ₂ O ₅ + packaging in boxes overwrapped with stretch PVC film | Delayed ripening | Joseph and Aworh, 1992 |
| papaya | Methyl jasmonateMJ at 10^{-5} M+ MAP (3–5 kPa O2 and 6–9 kPa CO2) at 10° C | 14-32 days | González- Aguilar <i>et al.</i> , 2003 |
| Peach | Ultrasound (40 kHz, 8.8 W/L, 10 min)+ salicylic acid (0.05 mM) | 6 days | Yang <i>et al.</i> , 2011 |
| | 38 °C for 12 h + 1 µmol L–1 Methyl jasmonate vapor at 20 °C for 24 h | Chilling injury suppressed for 3 weeks storage | Jin et al., 2009 |
| Pear | Gamma dose (1.5-1.7 kGy)+refrigerated storage (3±1 °C, RH 80%) | 8 days extension | Wani et al., 2008 |
| Plum | aqueous chlorine dioxide (40 mgL ⁻¹ ClO ₂ for 10 min) + ultrasound (100W for 10 min) | 60 days | Chen and Zhu, 2011 |
| | 0.5 μ L L-1 1-MCP at 0 °C for 24 h followed by storage at 10°C | 30 days | Minas <i>et al.</i> , 2013 |
| Pomegra- nate | Shrink wrapping+8°C storage temperature | 12 weeks | Nanda <i>et al.</i> , 2001 |
| Sapote | Brushing, Wax coating, 1-MCP treatment | 14 days | Ergun <i>et al.</i> , 2005 |
| Strawberry | Perforated LDPE (10 μ m thickness) +low temperature (0°C) | 15 days | Guerreiro <i>et al.</i> , 2013 |
| | UV-C light (1 kJ m ^{-2}), gaseous O3 (5000 mg L ^{-1}) and two active MAP conditions (superatmospheric O ₂ and CO ₂ -enriched atmospheres) | Delayed | Villalobos <i>et al.</i> , 2011 |
| | Calcium dip +chitosan coating | 4days-week for storage temperatures of 20 and 10°C | Hernández- Muñoz <i>et al.</i> , 2006, 2008 |
| Saskatoon fruit | Low temperature $(0.4^{\circ}C) + 2\% \text{ O2}$ | Quality maintained after 10 days | Rogiers and Knowles, 1998 |
| Tomato | 0.1 MPa pressure+ low temperature (13 °C) | Retention of firmness after 5 days | Liplap <i>et al.</i> , 2013a,b |
| Cherry tomato | 5–30 mg/l ozone gas for 0–20 min +MAP (6% $O_2/4\%$ CO_2), stored at 7°C | Salmonella Enteritidis died within 6 days | Das et al., 2006 |

Though the parameters discussed above has been effective in extending the shelf life of fresh fruits, when applied individually, their synergistic effect when combined with one or more parameters have been proved beneficial too. In the Table 4, some references on fresh whole fruits preservation using hurdle technology by employing two or more hurdles, is delineated.

Conclusions

Different hurdles are discussed vis-a vis extending the shelf life of fresh produce. Though more than 100 hurdles are practiced for food preservation; only a handful could be applied for the storage of fresh fruits. Milder doses of irradiation doses and storage at sub-optimal temperature would reduce the microbial load and inhibit ethylene biosynthesis; thus prolonging the shelf life. Application of organic acids, enzymes, chitin/chitosan, antimicrobial nisin, lactoferin, plant -derived antimicrobials, ozone, reuterin, electrolysed water, edible coating or MAP/CAP for the inhibition of other metabolic activities such as respiration would be suitable for climacteric fruits. Sanitizing agents for inhibiting Micro-flora could be chlorine water, intense light pulses, super high O2, N2O and noble gases. For

climacteric fruits care should be taken while choosing the packaging system. During the storage the fruits respire and leave out CO_2 and H_2O . This builds up CO_2 in the package, thus inducing ethylene biosynthesis. Ethylene production in large amount may cause uneven ripening of the fruits.

For non climacteric fruits, application of plant essential oils as anti microbial agent or any of the anti-

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microbial treatment such as irradiation, chemical treatment, application of O_3 , bacteriocins, essential oils, phenolics, microwave blanching, infrared and radio frequency heating would be sufficient, CAP, MAP, active packaging when stored at suboptimal temperature and relative humidity conditions can delay the respiration rate and senescence.

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